AMENDMENTS TO THE SPECIFICATION

Please amend paragraphs [0076], through [0079] and [0088] as follows:

[0076] PCR primers for the F gene:

Reference	Sequence	Position
MSF1	TGACCACGAGGTTACCTCTAC	(1057 matrix protein, forward) SEQ ID NO. 3
2FOV	TCCAAGTAGGTGGCACGCATA	(957, reverse) SEQ ID NO. 4
3FOV	AATTGACTACAGTATTCGGACC	(693, forward) SEQ ID NO. 5
4FOV	TGTTGACATTCCCAAGCTCAG	(1460, reverse) SEQ ID NO. 6
5FOV	GCTCAGTCATCGCTAACTGC	(1209, forward) SEQ ID NO. 7
6FOV	CGG AAT ATC AAG CGC CAT GTA	(168 of HN gene, reverse) SEQ ID NO. 8

[0077] Sequencing primers for F gene:

1FOV	TTAGAAAAAACACGGGTAGAA	(0, forward) SEQ ID NO. 9
7FOV	ACAGGACATTGACCACTTTGC	(300, forward) SEQ ID NO. 10
8FOV	CAGGTAACTCTACCTTCAGTCG	(902, forward) SEQ ID NO. 11
9FOV	CAACTCGATCAGTAATGCTTTGA	(1459, forward) SEQ ID NO. 12
10FOV	CCTAGATCAGATGAGAGCCACTACA	(1675, forward) SEQ ID NO. 13
11FOV	CTGCTGCATCTTCCCAACTG	(598, reverse) SEQ ID NO. 14
12FOV	GACTCTTGTATCCTACGGATAGA	(360, reverse) SEQ ID NO. 15
13FOV	GTACATACAGGCCGATGTATTGC	(1162, reverse) SEQ ID NO. 16
14FOV	AAGGTCTTTTGTTGCGCCTTTTG	(1653, reverse) SEQ ID NO. 17

[0078] PCR primers for the HN gene:

1HNOV	CGTTAGCCAAGTTGCGTTAGAG	(103, forward) SEQ ID NO. 18
2HNOV	CCGTCGAACCCTAACCTCC	(927, reverse) SEQ ID NO. 19
3HNOV	GTCTTGCAGTGTGAGTGCAAC	(799, forward) SEQ ID NO. 20
4HNOV	CCTCGCAAGGTGTGGTTTCTA	(1548, reverse) SEQ ID NO. 21
5HNOV	GCCACTCTTCATAGTCCTTATACA	(1397, forward) SEQ ID NO. 22
6HNOV	CCATGAGCTGTTTTGCCTTGTATCT	(intergenic HN/L, reverse)

(6HNOV) SEQ ID NO. 23

[0079] Sequencing primers for HN gene

7HNOV GCACCTATCCATGACCCAGATT (464, forward) SEQ ID NO. 24

8HNOV CGATACAATGACACATGCCCAGA (1106, forward) SEQ ID NO. 25

9HNOV GACCTATTGTCTCAGCATTGCTGA (1708, forward) SEQ ID NO. 26

10HNOV GGAACCAAGTGTAGATGTAATCT (319, reverse) SEQ ID NO. 27

11HNOV GAGGGTATTCGAGTGCAACCTGA (621, reverse) SEQ ID NO. 28

12HNOV GGTCTTCGCCTAAGGATGTTG (1247, reverse) SEQ ID NO. 29

13HNOV CTGAATTCTCCGAAGAGAGTAT (1761, reverse) SEQ ID NO. 30

14HNOV TGATCGCATGAGCACTGGCTG (1964, reverse) SEQ ID NO. 31

[0088] Each reaction mixture comprised of 25 µl RT-PCR Ready mix x2, 8 µl RNA, 5 µl of each forward and reverse sequencing primer and 7 µl DDH20. The components were well mixed and spun briefly prior to subjection to the RT-PCR reaction (48°C for 45 minutes for the RT reaction). Cycling parameters for the PCR were 94°C for 2 minutes (one cycle), 94°C (30 seconds), 60°C (1 minute) and 68°C (2 minutes) for 40 cycles and 68°C for 7 minutes. The PCR reaction mixes were loaded on 1% agarose gel and visualized using a UV Tran illuminator. Band size was estimated by comparing with DNA marker. DNA fragments were excised from the gel and purified using the Mini-elute Gel extraction kit (Qiagen). Each fragment was resuspended in ddH20. The DNAs were subjected to Sequencing analysis. The RT-PCR and sequencing primers were

NDV-1 T T G C A G C T G C A G G A A T T G T (4653 forward) SEQ ID NO. 32

NDV-2 C T A T A C A G T A T G A G G T G T C A A G (5540 reverse) SEQ ID NO. 33

NDV-4 G A A T T G A C T A C A G T A T T C G G (5189 FORWARD) SEQ ID NO. 34

NDV-5 G C G C G G T C C A T G A T T G A (6406 reverse) SEQ ID NO. 35